

Update on current applications of proteomic in the study of inflammatory bowel disease

Giulia Roda^a, Alessandra Caponi^a, Alessandro Sartini^a, Monica Cevenini^b, Carolina Colliva^c, Aldo Roda^c

S. Orsola-Malpighi Hospital, University of Bologna, Bologna, Italy

Abstract

Ulcerative colitis and Crohn's disease are relapsing and remitting chronic disorders. So far, endoscopy is the gold standard for their diagnosis, but less invasive diagnostic biomarkers are needed. Many authors have developed techniques to individuate biomarkers such as genetic testing factor or proteins in biological samples such as serum, plasma, and cellular subpopulations. A protein fingerprint pattern, patient-unique, specific for the diagnosis of inflammatory bowel disease (IBD) and potentially able to predict the future patterns of disease and to help in diagnosis, treatment, and prognosis is of increasing interest among researchers. Nowadays, a proteomic approach may be used in the identification of major alterations of proteins in IBD, but there is still a lack in the identification of a panel of biomarkers among a significant number of patients in large clinical trials. In this review, we analyze and report the current knowledge in proteomic application and strategies in the study of IBD.

Keywords Crohn's disease, ulcerative colitis, proteomic, biomarkers, metabolomic, nutrigenomic

Ann Gastroenterol 2012; 25 (4): 303-308

Introduction

Inflammatory bowel disease (IBD), i.e., ulcerative colitis (UC) and Crohn's disease (CD), are chronic inflammatory diseases, the pathogenesis of which is the result of altered immunological responses in a genetically susceptible host. Recently, the gut-associated microbial ecosystem has been explored as an important active player in IBD pathogenesis [1].

Nowadays, diagnosis of IBD is performed by endoscopic, histological, and radiographic tests, but even when performed by expert clinicians, they can result in diagnostic uncertainty. Moreover, early diagnosis is relevant in the therapeutic decision and thus in modifying the natural history of a disease.

None of the markers available have enough high sensitivity and specificity to allow an early and differential diagnosis between CD, UC and other colitis [2-38].

In the last decade, new technologies, such as genomics and proteomics, have combined in the study of IBD. The

knowledge of the genes involved in the etiopathogenesis of IBD and their products is of great interest for clinical use. Other -omic technologies, like metabolomics and nutrigenomics are emerging as new potential approaches in the discovery of new targets for the development of new drug therapies.

Many authors using new advanced methodologies such as proteomics but also metabolomics, nutrigenomics and subproteomics, have explored serological markers. The latter technique, besides investigating the proteome of different biological fluids, introduces the concept of functional proteomics and therefore might be of relevance because it focuses on the different cell compartment contents. Clinical subproteomics is achieving greater importance in the identification of IBD-related protein profiles or biomarkers. Moreover, new technologies for understanding the crosstalk between the microbiota and the host are emerging.

The aim of this review is to critically report the new bioanalytical tools in terms of suitability and real applicability to IBD biomarker discovery, reviewing the "state of the art" proteomic discovery biomarkers in IBD.

Proteomics

Analytical approaches

Mass spectrometry (MS) measures the mass to charge ratio (m/z) of ionized analytes, such as proteins or peptides. MS

^aGastroenterology Unit, S. Orsola-Malpighi Hospital (Giulia Roda, Alessandra Caponi, Alessandro Sartini); ^bDepartment of Clinical Medicine, University of Bologna, S. Orsola Hospital (Monica Cevenini); ^cDepartment of Pharmaceutical Sciences, University of Bologna (Carolina Colliva, Aldo Roda), Bologna, Italy

Conflict of Interest: None

Correspondence to: Prof. Aldo Roda, Department of Pharmaceutical Sciences, University of Bologna, Via Belmeloro 6, I-40126, Italy, e-mail: aldo.roda@unibo.it

Received 16 April 2012; accepted 15 May 2012

can be equipped with different ionization sources and mass analyzers. The most common are electro-spray ionization and matrix-assisted laser desorption ionization (MALDI) MS, while hybrid analyzer combining quadrupole and time-of-flight (TOF) are predominant [39]. Separation techniques play a pivotal role in the simplification of the sample. Proteins are separated on the basis of their molecular weight and isoelectric point using gel electrophoresis. It is the first protein separation technique that could be combined with MS. It allows cutting a single spot or band of the gel that is digested by proteases. This approach permits to perform qualitative and quantitative analysis at the protein level and to investigate post-translational modifications.

Liquid chromatography is another technique able to separate peptides prior to MS analysis. This procedure is faster and more reproducible than 2D-electrophoresis, it provides the identification of a larger amount of proteins. Quantitative analysis can be performed by a label-free approach or by labeling with isotopic peptides. Label-free quantitation is performed by comparison of the signals of the peptides eluted during the chromatographic separation. The performance of this approach can be affected by shifts in the retention time of the peptides between different runs. Isotopic labels can be distinguished among label strategy in enzymatic, metabolic, and chemical reactions. Enzymatic labeling can be performed during or after proteolytic digestion with proteases, while metabolic labeling can be applied to cell cultures by addition of isotope-enriched amino acids to the medium culture. Chemical label is the only label that can be applied to every type of biological sample [40-43]. These analytical approaches have been applied to a large variety of samples such as serum and plasma, freshly isolated cells, tissue and cell line in order to investigate the etiopathogenesis of IBD (Table 1).

Understanding the complexity of the data obtained by the analytical approaches described above is the goal of bioinformatics. Bioinformatics provides new algorithms to manage the large and heterogeneous amount of data such as new algorithms for image analysis of two-dimensional gels and for peptide mass fingerprinting and peptide fragmentation fingerprinting. Thus, bioinformatics emerges as an important approach to proteomic data sets, in order to understand the diversity between the normal and abnormal cell proteome of various biological systems.

Serum proteomics

Proteomics may contribute to biomarker discovery because it identifies a panel of proteins suitable as biomarkers in each biological sample [44-46].

Several groups have used serum for proteomic studies because of its simplicity. Meuwis first described several markers in IBD serum using surface-enhanced laser desorption/ionization /TOF MS such as platelet aggregation factor 4 (PAF4), myeloid-related protein 8, fibrinogen- α , and haptoglobin a2 (Hpa2). Moreover, they identified a correlation between PAF4 and the response to anti-tumor necrosis factor alpha

(anti-TNF- α) therapy. They demonstrated that anti-TNF- α non-responding CD patients have higher concentrations of PAF4 factor than healthy subjects [47,48]. Using different selective solid-phase bulk extraction, MALDI TOF MS and chemometric data analysis, we have found 20 proteins able to discriminate between healthy controls, CD and UC serum samples [49].

Cell proteomics

In normal condition, gut homeostasis derives from the cooperation of different populations of cells located throughout the mucosa and submucosa. From these interactions protein networks derive and create new interactions and cellular modifications.

Different proteomics approaches have been applied to *in vitro* models of IBD (colonic epithelial cell lines) as well as to cells derived from human samples and to different mouse models of colitis [50-52]. Hardwidge *et al* performed a large-scale proteomic study using liquid chromatography and MS to evaluate the response of colonic cell line to the enteropathogenic *Escherichia coli*. They described more than 2,000 proteins differently expressed in the presence or absence of the pathogen [52]. Several targets of inflammatory cytokines downstream pathway (tryptophanyl-tRNA synthetase, indoleamine-2,3-dioxygenase, heterogeneous nuclear ribonucleoprotein JKTBP, interferon-induced 35 kDa protein, proteasome subunit LMP2, and arginosuccinate synthetase) have been described in freshly isolated intestinal epithelial cells (IECs) samples from IBD patients. Overexpression of indoleamine-2,3-dioxygenase, an enzyme involved in tryptophan metabolism, was found in CD as well as in UC IECs [50,51].

Several proteins, like Annexin 2A, involved in cell death, signal transduction and energy metabolism, and induced as a response to stress, were found upregulated in inflamed area from UC IECs samples [53] and their increase correlated with inflammation and repair mechanisms. For example, protocadherins play a role in maintaining orderly growth during the re-epithelialization process, since they were found linked to the retention of the monolayer morphology of proliferating cells [54]. Hsieh *et al* using bidimensional electrophoresis (2DE) and MS, identified differences in the expression of proteins among active and inactive UC such as mitochondrial proteins, proteins involved in energy generation, or stress-response proteins [55]. Because of the need for information concerning the pathogenesis of IBD, the variations in the protein expression of lymphocytes and mucosa were studied in rats with induced colitis and CD and UC patients. Recently, the protein profile of lymphocytes from an antigen-specific model of colitis has been performed using proteomic approaches. Liu *et al* identified 26 altered proteins in lymphocytes isolated from rat with 2,4,6-trinitrobenzene sulfonic acid (TNBS)-induced colitis. Among these proteins some are involved in inflammation, such as myeloid-related protein 14, a potential mediator of p38 mitogen-activated protein kinases -dependent functional responses, apoptosis,

metabolism, such as ATP-citrate synthase, regulation of cell cycle, cell proliferation, signal transduction, such as nucleoside diphosphate kinases, and ubiquitin conjugating enzyme E2N [56].

Direct tissue analysis

Histology-directed protein profiling allows acquiring spectra from areas of mucosa or submucosa within a single tissue section without preparation of a sample.

Berndt's group performed a proteomic analysis of inflamed and uninfamed areas of the gut from IBD patients using a novel automated multidimensional, fluorescence-based microscopy robot technology demonstrating that IBD has a greater number of CDA β CD7-memory T cells [57].

Recently, other groups have used the ability of MALDI to perform histology-directed cellular protein analysis of tissues. M'Koma analyzed mucosal and submucosal layers of CD and UC colon resection samples after histologic assessment. MALDI-MS appeared to be capable to distinguish CD and UC while profiling the colonic submucosa [58].

Subproteomics

The biological fluids and cells are very complex samples composed of thousands of peptides, proteins, and products of metabolism. Subproteomic is crucial in determining a protein profile of a single subset of cells or a single compartment by applying new strategies for the fractionation, separation, and enrichment of proteins derived from a specific compartment or cell subtype.

Examples of subproteomes that are being studied include large macromolecular complexes and cellular machines, specific classes of proteins and organelles. Although the same analytical technologies may usually be employed for global and targeted proteomic studies, the latter studies require specific initial strategies to isolate the appropriate subproteome components.

Subcellular proteomics

Huber *et al* introduced the concept of subcellular proteomics. Subcellular fractionation is a flexible approach that reduces sample complexity and is efficiently combined both with high-resolution 2D gel/MS analysis and with gel-free independent techniques [59]. Subcellular fractionation allows access to intracellular organelles and multiprotein complexes and tracking proteins that shuttle between different compartments (e.g., between the cytoplasm and nucleus) [60]. Subproteomics represents a relevant approach to the study of IBD where there are defects in different compartments. Rectal biopsy specimens from control subjects and from patients with CD, non-rectal CD, and acute UC were subjected to sucrose density gradient centrifugation. The activity of enzymes such

as 5'nucleotidase (plasma membrane), malate dehydrogenase (mitochondria), catalase (peroxisomes), lactate dehydrogenase (cytosol), N-acetyl-beta-glucosaminidase (lysosomes), neutral-alpha-glucosidase (endoplasmic reticulum), were tested [61].

We evaluated the proteome of the subcellular fractions (nuclei, membranes, and cytosols) of IECs isolated from healthy subjects and CD, using a label-free liquid chromatography-MS approach. We found in CD an increase in proteins such as heat shock 70 kDa protein 5, trypsin alpha-1 precursor, and proteins whose upregulation can be explained by the increased activity of IECs in the inflammatory state. A lower abundance in CD of proteins such as Annexin A1, a mediator of the antiinflammatory actions of glucocorticoids, and malate dehydrogenase was found too [62].

Serum subproteomics: microparticles

Recent studies have focused on the relationship between inflammation and blood coagulation in the pathogenesis of IBD, where there is an increase in the number of circulating platelets.

In CD, platelet activation is demonstrated by positivity for P-selectin (CD62P), b-thromboglobulin, and platelet products such as platelet factor 4 (PF4) and CD40 ligand (CD40L). Activated platelets produce platelet-derived microparticles (PDMPs) in response to different stimuli. PDMPs range in size from 0.02 to 0.5 μ m and carry several antigenic characteristics of intact platelets such as glycoprotein (GP) IIb/IIIa, GPIb/IX, and P-selectin (CD62P). The generation of microparticles can be the consequence of proinflammatory cytokines production [63-69].

The PDMPs are membrane-derived microvesicles (MVs) released from the cell surface and implicated in cell-to-cell communication. MVs may represent vehicles that can facilitate interaction between target cells by surface expressed ligands; they can transfer surface receptor, deliver proteins, mRNA, intact organelles, and modulate the cell-to-cell network. Huber's group investigated the role of microvesicles secreted from colon cancer cells in inducing T cell apoptosis and escaping the immune system. They demonstrated that tumor microparticles are able to induce the apoptosis of activated CD8 β T cells through the expression of both a granular pattern of tumor necrosis factor-related apoptosis-inducing ligand and the Fas ligand [70].

Our group has speculated that PDMPs from CD patients contain specific esopeptidase which involves the fibrinopeptide as substrate [71].

From metabolomics to nutrigenomics

Metabolomics is the "systematic study of the unique chemical fingerprints that specific cellular processes leave behind"-specifically, the study of their small-molecule metabolite profiles. The metabolome represents the collection of all metabolites in a biological organism, which are the

end products of its gene expression. Metabolic profiling can give an instantaneous snapshot of the physiology and pathophysiology of that cell.

The urinary metabolomic profiles in interleukin 10 gene-deficient mice using high performance liquid chromatography - MS revealed that changes in trimethylamine and fucose correlates with disease activity [72]. Marchesi characterized fecal extracts using a noninvasive metabolomics approach, which combines high-resolution ¹H NMR spectroscopy and multivariate pattern recognition techniques. The fecal extracts from IBD patients were characterized by reduced levels of butyrate, acetate, methylamine, and trimethylamine in comparison with a control population, suggesting changes in the gut microbial community. Elevated quantities of amino acids were present in the feces from both disease groups. The roles that amino acids play in immunity and inflammation are well defined, and the relationship between IBD and certain amino acids has recently been studied [73]. Ooi *et al* studied the levels of amino acids and trichloroacetic acid (TCA) cycle-related molecules in the colonic tissues and sera of patients with UC by gas chromatography MS. They observed lower levels of 16 amino acids and 5 TCA cycle-related molecules in UC patients than in CD patients and healthy subjects [74].

The role of bacteria in the pathogenesis of several gastrointestinal disorders is well established. Kumari first introduced the concept of quorum sensing in IBD. Altered bacterial balance in the gut may lead to inflammation. Bacteria create communications among themselves (quorum sensing) to maintain their behavior and colonization. Quorum sensing enables bacteria to communicate with each other and control expression of certain specialized genes by producing and responding to extracellular signals in proportion to cell density.

Kumari proposed a method based on genetically engineered whole-cell sensing systems to detect quorum-sensing molecules in biological samples and to use these molecules as biomarkers [75].

Nutrigenomics is defined as “a transdisciplinary approach to understand the subtle but contentious impact of nutrition as prime environmental trigger in shaping the dynamic range between health and disease”. Proteomic analysis of primary intestinal epithelial cells, as mentioned above, has confirmed a network of differentially regulated proteins clustered around mitochondrial stress mechanisms. Different authors have shown a connection between mitochondrion and endoplasmic reticulum in triggering stress-associated unfolded protein response that could implement inflammation. Nutrigenomic together with proteomic and genomic could represent new progress in understanding the network between diet-gene and/or diet-microbe interaction and to discover new links between metabolome and IBD pathogenesis [76].

Conclusions

In this review we have summarized the current “state of the art” in emerging proteomic technologies and their applications

in the study of IBD. Despite the fact that new advanced tools are available, few groups have attempted to apply proteomic study for the discovery of new biomarkers in IBD.

The complex etiopathogenesis of CD and UC which are the result of modifications occurring at different levels may be revealed by the study of protein modification using proteomic and the related technologies. Moreover, because several cell subpopulations, compartments are affected, subproteomics may allow target proteomic studies.

Furthermore, the emerging role of microbiota in maintaining the intestinal homeostasis point out the need to integrate proteomics together with metabolomics and nutrigenomics. Thus, understanding the diet-host-microbiota interactions may help to identify novel targets in the etiopathogenesis of IBD.

References

1. Abraham C, Medzhitov R. Interactions between the host innate immune system and microbes in inflammatory bowel disease. *Gastroenterology* 2011;**140**:1729-1737.
2. Dotan I. New serologic markers for inflammatory bowel disease diagnosis. *Dig Dis* 2010;**28**:418-423.
3. Lewis JD. The utility of biomarkers in the diagnosis and therapy of inflammatory bowel disease. *Gastroenterology* 2011;**140**:1817-1826.e2.
4. Beniwal P, Harrell L. The status of diagnostic markers for inflammatory bowel disease. *Curr Gastroenterol Rep* 2010;**12**:479-484.
5. Beaven SW, Abreu MT. Biomarkers in inflammatory bowel disease. *Curr Opin Gastroenterol* 2004;**20**:318-327.
6. Price A. Overlap in the spectrum of non-specific inflammatory bowel disease-colitis indeterminate. *J Clin Pathol* 1978;**31**:567-577.
7. Joossens S, Reinisch W, Vermeire S, et al. The value of serologic markers in indeterminate colitis: a prospective follow-up study. *Gastroenterology* 2002;**122**:1242-1247.
8. Vermeire S, Van Assche G, Rutgeerts P. Laboratory markers in IBD: useful, magic or unnecessary toys? *Gut* 2006;**55**:426-431.
9. Poullis AP, Zar S, Sundaram KK, et al. A new, highly sensitive assay for C-reactive protein can aid the differentiation on inflammatory bowel disorders from constipation and diarrhoea predominant functional bowel disorders. *Eur J Gastroenterol Hepatol* 2002;**14**:409-412.
10. Koelewijn CL, Schwartz MP, Samsom M, et al. C-reactive protein levels during a relapse of Crohn's disease are associated with the clinical course of the disease. *World J Gastroenterol* 2008;**14**:85-89.
11. Saxon A, Shanahan F, Landers C, et al. A distinct subset of antineutrophil cytoplasmic antibodies is associated with inflammatory bowel disease. *J Allergy Clin Immunol* 1990;**86**:202-210.
12. Eggena M, Targan SR, Iwanczyk L, et al. Phage display cloning and characterization of an immunogenetic marker (perinuclear anti-neutrophil cytoplasmic antibody) in ulcerative colitis. *J Immunol* 1996;**156**:4005-4011.
13. Eggena M, Cohavy O, Parseghian MH, et al. Identification of histone H1 as a cognate antigen of the ulcerative colitis-associated marker antibody pANCA. *J Autoimmun* 2000;**14**:83-97.
14. Vecchi M, Sinico A, Bianchi MB, et al. Recognition of bactericidal/permeability-increasing protein by perinuclear anti-neutrophil cytoplasmic antibody-positive sera from ulcerative colitis patients:

- prevalence and clinical significance. *Scand J Gastroenterol* 1998;**33**:1284-1288.
15. Papp M, Norman GL, Altorjay I, et al. Utility of serological markers in inflammatory bowel diseases: gadget or magic? *World J Gastroenterol* 2007;**13**:2028-2036.
 16. Mainardi E, Villanacci V, Bassotti G, et al. Diagnostic value of serological assays in pediatric inflammatory bowel disorders. *Digestion* 2007;**75**:210-214.
 17. Foley KF, Kao P. Biomarkers for inflammatory bowel disease. *Clin Lab Sci* 2007;**20**:84-88.
 18. Joossens S, Colombel JF, Vermeire S, et al. Panel of serologic antibodies in patients with indeterminate colitis. *Gastroenterology* 2003;**124**:A323.
 19. Vasiliauskas EA, Kam LY, Karp LC, et al. Marker antibody expression stratifies Crohn's disease into immunologically homogeneous subgroups with distinct clinical characteristics. *Gut* 2000;**47**:487-496.
 20. Vermeire S, Peeters M, Vlietinck R, et al. Anti-Saccharomyces cerevisiae antibodies (ASCA), phenotypes of IBD, and intestinal permeability: a study in IBD families. *Inflamm Bowel Dis* 2001;**7**:8-15.
 21. Vasiliauskas EA, Plevy SE, Landers CJ, et al. Perinuclear antineutrophil cytoplasmic antibodies in patients with Crohn's disease define a clinical subgroup. *Gastroenterology* 1996;**110**:1810-1819.
 22. Papp M, Altorjay I, Dotan N, et al. New serological markers for inflammatory bowel disease are associated with earlier age at onset, complicated disease behaviour, risk for surgery, and NOD2/CARD15 genotype in a Hungarian IBD cohort. *Am J Gastroenterol* 2008;**103**:665-681.
 23. Kim BC, Park S, Han J, et al. Clinical significance of anti-Saccharomyces cerevisiae antibody (ASCA) in Korean patients with Crohn's disease and its relationship to the disease clinical course. *Dig Liver Dis* 2007;**39**:610-616.
 24. Ferrante M, Henckaerts L, Joossens M, et al. New serological markers in inflammatory bowel disease are associated with complicated disease behaviour. *Gut* 2007;**56**:1394-1403.
 25. Klebl FH, Bataille F, Bertea CR, et al. Association of perinuclear anti-neutrophil cytoplasmic antibodies and anti-Saccharomyces cerevisiae antibodies with Vienna classification subtypes of Crohn's disease. *Inflamm Bowel Dis* 2003;**9**:302-307.
 26. Sandborn WJ, Landers CJ, Tremaine WJ, et al. Association of anti-neutrophil cytoplasmic antibodies with resistance to treatment of left-sided ulcerative colitis: results of a pilot study. *Mayo Clin Proc* 1996;**71**:431-436.
 27. Taylor KD, Plevy SE, Yang H, et al. ANCA pattern and LTA haplotype relationship to clinical responses to anti-TNF antibody treatment in Crohn's disease. *Gastroenterology* 2001;**120**:1347-1355.
 28. Barahona-Garrido J, Camacho-Escobedo J, García-Martínez CI, Tocay H, Cabiedes J, Yamamoto-Furusho JK. Antinuclear antibodies: a marker associated with steroid dependence in patients with ulcerative colitis. *Inflamm Bowel Dis* 2009;**15**:1039-1043.
 29. Angriman I, Scarpa M, D'Inca R, et al. Enzymes in feces: useful markers of chronic inflammatory bowel disease. *Clin Chim Acta* 2007;**381**:63-68.
 30. Sutherland AD, Geary RB, Frizelle FA. Review of fecal biomarkers in inflammatory bowel disease. *Dis Colon Rectum* 2008;**51**:1283-1291.
 31. Xiang JY, Ouyang Q, Li GD, et al. Clinical value of fecal calprotectin in determining disease activity of ulcerative colitis. *World J Gastroenterol* 2008;**14**:53-57.
 32. Peterson CG, Sangfelt P, Wagner M, et al. Fecal levels of leukocyte markers reflect disease activity in patients with ulcerative colitis. *Scand J Clin Lab Invest* 2007;**67**:810-820.
 33. Shitrit AB, Braverman D, Stankiewicz H, et al. Fecal calprotectin as a predictor of abnormal colonic histology. *Dis Colon Rectum* 2007;**50**:2188-2193.
 34. Bunn SK, Bisset WM, Main MJ, et al. Fecal calprotectin as a measure of disease activity in childhood inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2001;**32**:171-177.
 35. Sipponem T, Savilahti E, Kärkkäinen P, et al. Fecal calprotectin, lactoferrin, and endoscopic disease activity in monitoring anti-TNF-alpha therapy for Crohn's disease. *Inflamm Bowel Dis* 2008;**14**:1392-1398.
 36. Walker TR, Land ML, Kartashov A, et al. Fecal lactoferrin is a sensitive and specific marker of disease activity in children and young adults with inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2007;**44**:414-422.
 37. Bunn SK, Bisset WM, Main MJ, et al. Fecal calprotectin: validation as a noninvasive measure of bowel inflammation in childhood inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2001;**33**:14-22.
 38. Bjarnason I, Sherwood R. Fecal calprotectin: a significant step in the non-invasive assessment of intestinal inflammation. *J Pediatr Gastroenterol Nutr* 2001;**33**:11-13.
 39. Aebersold R, Mann M. Mass spectrometry-based proteomics. *Nature* 2003;**422**:198-207.
 40. Wittmann-Liebold B, Graack HR, Pohl T. Two dimensional gel electrophoresis as tool for proteomics studies in combination with protein identification by mass spectrometry. *Proteomics* 2006;**6**:4688-4703.
 41. Bantscheff M, Schirle M, Sweetman G, et al. Quantitative mass spectrometry in proteomics: a critical review. *Anal Bioanal Chem* 2007;**389**:1017-1031.
 42. Iliuk A, Galan J, Tao WA. Playing tag with quantitative proteomics. *Anal Bioanal Chem* 2009;**393**:503-513.
 43. Gygi SP, Rist B, Gerber SA, et al. Quantitative analysis of complex protein mixtures using isotope-coded affinity tags. *Nat Biotechnol* 1999;**17**:994-999.
 44. Apweiler R. Approaching clinical proteomics: current state and future fields of application in fluid proteomics. *Clin Chem Lab Med* 2009;**47**:724-744.
 45. Li X, Conklin L, Alex P. New serological biomarkers of inflammatory bowel disease. *World J Gastroenterol* 2008;**14**:5115-5124.
 46. Alex P, Gucek M, Li X. Applications of proteomics in the study of inflammatory bowel disease: current status and future directions with available technologies. *Inflamm Bowel Dis* 2008;**4**:616-629.
 47. Meuwis MA, Fillet M, Geurts P, et al. Biomarker discovery for inflammatory bowel disease, using proteomic serum profiling. *Biochem Pharmacol* 2007;**73**:1422-1433.
 48. Meuwis MA, Fillet M, Lutteri L, et al. Proteomics for prediction and characterization of response to infliximab in Crohn's disease: a pilot study. *Clin Biochem* 2008;**41**:960-967.
 49. Nanni P, Parisi D, Roda G, et al. Serum protein profiling in patients with inflammatory bowel diseases using selective solid-phase bulk extraction, matrix-assisted laser desorption/ionization time-of-flight mass spectrometry and chemometric data analysis. *Rapid Commun Mass Spectrom* 2007;**21**:4142-4148.
 50. Barcelo-Batllo S, André M, Servis C, et al. Proteomic analysis of cytokine induced proteins in human intestinal epithelial cells: implications for inflammatory bowel diseases. *Proteomics* 2002;**2**:551-560.
 51. Felley-Bosco E, André M. Proteomics and chronic inflammatory bowel diseases. *Pathol Res Pract* 2004;**200**:129-133.
 52. Hardwidge P, Rodriguez-Escudero I, Goode D, et al. Proteomic analysis of the intestinal epithelial cell response to enteropathogenic Escherichia coli. *J Biol Chem* 2004;**279**:20127-20136.
 53. Shkoda A, Werner T, Daniel H, et al. Differential protein expression profile in the intestinal epithelium from patients with inflammatory bowel disease. *J Proteome Res* 2007;**6**:1114-1125.

54. Fogt F, Jian BO, Krieg RC, et al. Proteomic analysis of mucosal preparations from patients with ulcerative colitis. *Mol Med Reports* 2008;**1**:51-54.
55. Hsieh SY, Shih TC, Yeh CY, et al. Comparative proteomic studies on the pathogenesis of human ulcerative colitis. *Proteomics* 2006;**6**:5322-5331.
56. Liu BG, Cao YB, Cao YY. Altered protein profile of lymphocytes in an antigen-specific model of colitis: a comparative proteomic study. *Inflamm Res* 2007;**56**:377-384.
57. Berndt U, Bartsch S, Philipsen L, et al. Proteomic analysis of the inflamed intestinal mucosa reveals distinctive immune response profiles in Crohn's disease and ulcerative colitis. *J Immunol* 2007;**179**:295-304.
58. M'Koma AE, Seeley EH, Washington MK, et al. Proteomic profiling of mucosal and submucosal colonic tissues yields protein signatures that differentiate the inflammatory colitides. *Inflamm Bowel Dis* 2011;**17**:875-883.
59. Huber LA, Pfaller K, Vietor I. Organelle proteomics: implications for subcellular fractionation in proteomics. *Circ Res* 2003;**92**:962-968.
60. Groen AJ, de Vries SC, Lilley KS. A proteomics approach to membrane trafficking. *Plant Physiol* 2008;**147**:1584-1589.
61. O'Morain C, Smethurst P, Levi J, et al. Subcellular fractionation of rectal biopsy homogenates from patients with inflammatory bowel disease. *Scand J Gastroenterol* 1985;**20**:209-214.
62. Nanni P, Mezzanotte L, Roda G, et al. Differential proteomic analysis of HT29 Cl.16E and intestinal epithelial cells by LC ESI/QTOF mass spectrometry. *J Proteomics* 2009;**72**:865-873.
63. Andoh A, Yoshida T, Yagi Y, et al. Increased aggregation response of platelets in patients with inflammatory bowel disease. *J Gastroenterol* 2006;**41**:47-54.
64. Chamouard P, Desprez D, Hugel B, et al. Circulating cell-derived microparticles in Crohn's disease. *Dig Dis Sci* 2005;**50**:574-580.
65. Andoh A, Tsujikawa T, Hata K, et al. Elevated circulating platelet-derived microparticles in patients with active inflammatory bowel disease. *Am J Gastroenterol* 2005;**100**:2042-2048.
66. Srirajaskanthan R, Winter M, Muller AF. Venous thrombosis in inflammatory bowel disease. *Eur J Gastroenterol Hepatol* 2005;**17**:697-700.
67. Choi SK, Kasturi L, Tavakoly A, Safier HL. Ulcerative colitis diagnosed in a patient with venous thromboembolism. *Hosp Physician* 2008;**44**:32-36.
68. Irving PM, Pasi KJ, Rampton DS. Thrombosis and inflammatory bowel disease. *Clin Gastroenterol Hepatol* 2005;**3**:617-628.
69. Pap E, Pállinger E, Pásztói M, Falus A. Highlights of a new type of intercellular communication: microvesicle-based information transfer. *Inflamm Res* 2009;**58**:1-8.
70. Huber V, Fais S, Iero M, et al. Human colorectal cancer cells induce T-cell death through release of proapoptotic microvesicles: role in immune escape. *Gastroenterology* 2005;**128**:1796-1804.
71. Nanni P, Levander F, Roda G. A label-free nano-liquid chromatography-mass spectrometry approach for quantitative serum peptidomics in Crohn's disease patients. *J Chromatogr B Analyt Technol Biomed life Sci* 2009;**877**:3127-3136.
72. Murdoch TB, Fu H, Farlane SM, et al. Urinary metabolic profiles of inflammatory bowel disease in interleukin-10 gene-deficient mice. *Anal Chem* 2008;**80**:5524-5531.
73. Marchesi JR, Holmes E, Khan F, et al. Rapid and noninvasive metabolomic characterization of inflammatory bowel disease. *J Proteome Res* 2007;**6**:546-551.
74. Ooi M, Nishiumi S, Yoshie T, et al. GC/MS-based profiling of amino acids and TCA cycle-related molecules in ulcerative colitis. *Inflamm Res* 2011;**60**:831-40.
75. Kumari A, Pasini P, Daunert S. Detection of bacterial quorum sensing N-acyl homoserine lactones in clinical samples. *Anal Bioanal Chem* 2008;**391**:1619-1627.
76. Haller D. Nutrigenomics and IBD: the intestinal microbiota at the cross-road between inflammation and metabolism. *J Clin Gastroenterol* 2010;**44** (Suppl 1):S6-S9.